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## A New Valence Tautomerism: Thermal Rearrangement of cis-2-Vinyl-3-ethynyl Three-Membered Heterocycles

Sir:

In recent years, there has been a considerable interest in the Cope rearrangement of 2,3-divinyl<sup>1</sup> and 2,3-diethynyl<sup>2</sup> three-membered rings. Our continuing interest in the thermolytic behavior of 1,5-enynes<sup>3a,b</sup> and the recently published rearrangement of cis-1-ethynyl-2-vinylcyclopropane,3c prompt us to report on our study of the valence isomerization of cis-1-ethynyl-2-vinyloxirane (1a) and cis-N-tert-butyl-2-ethynyl-3-vinylaziridine (1b).

The desired starting material **1a** was prepared by treatment of 3,4-dihydroxy-1,5-hexenyne<sup>4</sup> (erythro + threo) with 2 equiv of sodium hydride, and 1 equiv of p-toluenesulfonyl chloride in ether. A mixture of cis- and trans-1a was obtained (52% yield, cis:trans 1:0.7) and separated by preparative vapor phase chromatography. Deuterated 1c was prepared by stirring 1a with BaO in a large excess of D<sub>2</sub>O.<sup>5</sup> Aziridine 1b was prepared conveniently by aminolysis of *cis*-1a (46% yield), followed by cyclization of the intermediate three amino alcohol<sup>6</sup> with  $Ph_3PCl_2$  at room temperature<sup>7</sup> (31% yield). The structures of **1a,b** were established by NMR spectroscopy.<sup>8</sup>

• Bi

1b

Thermal rearrangements were conducted in sealed tubes in inert solvents ( $C_6H_6$ ,  $CCl_4$ ) over the temperature range 80–130 °C. These reactions gave rise to a single product: cis-1-carboxaldehyde-2-ethynylcyclopropane (3a) and N-tert-butyl-1H-azepine (4), respectively, from cis-1a and cis-1b. The structure of cis-3a was established by its straightforward spectral characteristics: <sup>1</sup>H NMR (60 MHz, C<sub>6</sub>H<sub>6</sub>,  $\delta_{MeaSi}$ )  $\dot{O}=5-4\nabla_6^3-2^{2}=19.22$  (m, 1 H, H<sub>5</sub>), 2.02 (d, 1 H, J = 1.6 Hz, H<sub>1</sub>), 1.85-1.50 (m, 2 H, H<sub>4</sub> and H<sub>3</sub>) 1.46-0.8 (m, 2 H, H<sub>6</sub>); <sup>13</sup>C NMR (15.08 MHz, CDCl<sub>3</sub>,  $\delta_{Me4Si}$ ) 69.1 (d, C<sub>1</sub>), 81.0 (d, C<sub>2</sub>), 8.7 (d, C<sub>3</sub>), 27.7 (d, C<sub>4</sub>), 200.7 (d, C<sub>5</sub>), 14.4 (t, C<sub>6</sub>); IR (CHCl<sub>3</sub>, cm<sup>-1</sup>), 3200, 2100, 1705; MS (70 eV, m/e, rel intensity %) 94 (M<sup>+</sup>, 5), 65 (100), as well as by its facile conversion to *trans*-3a<sup>9</sup> by thermolysis in a flow system<sup>10</sup> at 350 °C.

4

The structural assignment of 4 was based on its <sup>1</sup>H NMR spectrum (60 MHz, CCl<sub>4</sub>,  $\delta_{Me_4Si}$ ) 5.87 (t, 2 H, H-C<sub>4</sub>), 5.30  $(d, 2 H, J = 7.5 Hz, H-C_2) 4.95 (2t, 2 H, H-C_3), 1.12 (s, 9 H, H-C_3)$ t-Bu); the ethylenic part of the spectrum is very similar to that of N-carbalkoxy-1 $\hat{H}$ -azepines.<sup>11</sup> The <sup>13</sup>C NMR spectrum (15.08 MHz, CDCl<sub>3</sub>,  $\delta_{Me_4Si}$ ) 26.5 (methyls) 52.4 (quater, C) 114.4, 132.1, and 135.8 (C2, C3, and C4) confirmed this structural assignment.

The rearrangement of 1a is stereospecific and follows a clean first-order rate law<sup>12</sup> (up to 70% reaction) with respect to starting material. The calculated rate constants ( $\times 10^3$  mn) were determined by least-squares analysis of the experimental data:  $k(102^{\circ}.8) = 2.70, k(110^{\circ}.5) = 5.64, k(113^{\circ}.6) = 7.83,$  $k(116^{\circ}8) = 10.58, k(120^{\circ}6) = 14.61, k(130^{\circ}6) = 29.10$ . The activation parameters ( $\Delta H^{\pm} = 25.1 \pm 1.7 \text{ kcal mol}^{-1}, \Delta S^{\pm} =$  $-3 \pm 3$  eu) are compatible with a Cope rearrangement. The enthalpy of activation for this rearrangement is only 2.4 kcal mol<sup>-1</sup> higher than that for the rearrangement of cis-divinyloxirane.1i

The following mechanism (Scheme II) is proposed to ac-

Scheme II



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count for this new thermal isomerization. In the first step, a highly strained seven-membered heterocycle 2a is formed via a [3,3] sigmatropic rearrangement of 1a. This species may either give 3a or return to 1a by means of [3,3]sigmatropic shifts. Since the estimated heat of formation of 3a (calculated by the method of Benson et al.<sup>13</sup>) is  $\sim 19$  kcal mol<sup>-1</sup> less than that of 1a, the reaction proceeds in the expected direction, i.e.,  $1a \rightarrow$ **3a.** In contrast to the isomerization of *cis*-1-ethynyl-2-vinylcyclopropane,<sup>3c</sup> no dimers<sup>14</sup> were formed from the allenic intermediate 2a. The above mechanism is supported by the analogous conversion of deuterated compound 1c to 3c. The structure of 3c is confirmed by NMR: the spectrum reveals only one cyclopropane hydrogen at  $\delta$  1.85–1.50; moreover, the signal of the acetylenic hydrogen appears as a singlet.

Thermal rearrangement of 1b to 3b should also occur since the heat of formation of **3b** is estimated<sup>13</sup> to be less than  $\sim 9$ kcal  $mol^{-1}$  that of **1b**. Nevertheless, only the formation of **4** is observed when 1b is heated at 90 °C for 20 min. A pathway consistent with this fact would be a 1,3-hydrogen shift from the proposed intermediate 2b. Since a thermal concerted 1,3-shift is forbidden by the Woodward-Hoffman rules,<sup>15</sup> we suggest that the hydrogen transfer occurs intramolecularly and is catalyzed by the nitrogen atom in **2b**: one of the two allylic hydrogens is near the nitrogen atom, because the six centered transition state leading to 2b must generate a cis double bond. This hypothesis for the formation of the intermediate **2b** is further supported by the fact that 4 is the only product formed when cis- 3b<sup>16</sup> is subjected to flow pyrolysis<sup>10</sup> at 350 °C.<sup>17</sup>

It may be asked why different pathways are observed when 1a and 1b are submitted to pyrolysis. This can be attributed to the higher basicity of the nitrogen atom over the oxygen atom.

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(8) cis-1a: <sup>1</sup>H NMR (60 MHz, CCl<sub>4</sub>,  $\delta_{Me_4Si}$ ) 2.32 (d, 1 H, J = 1.6 Hz, H<sub>1</sub>), - <sup>5</sup>--- 4 

3.26–3.50 (m, 2 H, H<sub>3</sub> and H<sub>4</sub>), 5.16–6.00 (m, 3 H, H<sub>5</sub> and H<sub>6</sub>). <sup>13</sup>C NMR (15.08 MHz, CDCl<sub>3</sub>,  $\delta_{Me_4Si}$ ) 74.1 (d, C<sub>1</sub>), 78.6 (d, C<sub>2</sub>), 46.0 (d, C<sub>3</sub>), 57.9 (d, C<sub>4</sub>), 132.6 (d, C<sub>5</sub>), 122.3 (t, C<sub>6</sub>). MS (70 eV, *m/e*, rel intensity %) 94 (M<sup>+</sup>, 5), 65 (100). cis-1b: <sup>1</sup>H NMR (60 MHz, CCl<sub>4</sub>,  $\delta_{Me_4Si}$ ) 0.98 (s, 9 H, H<sub>8</sub>), 1.88



(m, 1 H, H<sub>5</sub>), 2.25 (m, 2 H, H<sub>1</sub> and H<sub>3</sub>), 5.01–5.70 (m, 3 H, H<sub>5</sub> and H<sub>6</sub>).  $^{13}C$ NMR (15.08 MHz, CDCl<sub>3</sub>,  $\delta_{Me_4Sl}$ ) 69.0 (d, C<sub>1</sub>) 82.0 (d, C<sub>2</sub>), 26.9 (d, C<sub>3</sub>), 39.9 (d, C<sub>4</sub>), 136.2 (d, C<sub>5</sub>), 118.0 (t, C<sub>6</sub>), 54.1 (s, C<sub>7</sub>), 26.3 (q, C<sub>8</sub>). MS (70 eV, *m*/e, rel intensity %) 149 (M<sup>+</sup>, 32), 93 (100).

- (9) Spectral data for *trans*-3a; <sup>1</sup>H NMR (60 MHz, C<sub>6</sub>H<sub>6</sub>,  $\delta_{Me_4Si}$ ) 8.96 (d, 1 H, J = 3.8 Hz, H<sub>5</sub>), 2.10–1.36 (m, 2 H, H<sub>3</sub> and H<sub>4</sub>) 1.88 (d, 1 H, J = 1.6 Hz, H<sub>1</sub>), 1.30-0.50 (m, 2 H, H<sub>6</sub>). MS (70 eV, m/e, rel intensity %) 94 (M<sup>+</sup>, 4), 65 (100)
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Scheme III



The authors would like to thank a referee for this suggestion.

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$$(^{\circ}CH_3)_3C - N = {}^{\circ} - {}^{\circ} = {}^{\circ}$$

Hz, H<sub>5</sub>), 2.25–1.60 (m, 2 H, H<sub>3</sub> and H<sub>4</sub>) 1.95 (d, 1 H, H 1.8 Hz, H<sub>1</sub>), 1.53–0.82 (m, 2 H, H<sub>6</sub>), 1.20 (s, 9 H, H<sub>8</sub>). <sup>13</sup>C NMR (15.08 MHz, CDCl<sub>3</sub>,  $\delta_{Me_4Sl}$ ) 67.4 (d, C<sub>1</sub>), 82.9 (d, C<sub>2</sub>), 6.9 (d, C<sub>3</sub>), 22.7 (d, C<sub>4</sub>), 158.6 (d, C<sub>5</sub>), 14.3 (t, C<sub>6</sub>), 57.1 (s, C<sub>7</sub>), 29.8 (q, <sub>8</sub>). MS (70 eV, *m/e*, rel intensity %) 149 (M<sup>+</sup>, 28), 93 (100).

(17) Compound 3b is stable under the milder conditions (90 °C) used for rearrangement of 1b.

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## The Crystal Structure of the Mushroom Toxin $\beta$ -Amanitin<sup>1</sup>

Sir:

The deadly poisonous mushroom Amanita phalloides contains a number of cyclic peptides which can be classified as phallotoxins (heptapeptides), amatoxins (octapeptides), and antamanide, a decapeptide antagonist of the phallotoxins. The amatoxins cause death by destroying liver cells and damaging the secretory cells of the convoluted tubules in the kidney via inhibition of RNA polymerase II.<sup>2,3</sup> Although the chemical sequences of these cyclopeptides have been determined, only antamanide has been subjected to a three-dimensional structure analysis.4

We wish to report the x-ray crystallographic structure determination of the amatoxin  $\beta$ -amanitin, isolated and purified from American Amanita phalloides.<sup>5</sup>  $\beta$ -Amanitin (1),  $C_{39}H_{53}SO_{15}N_{9}$ , has the chemical sequence cyclo (L- $\alpha$ -aspartyl-4-hydroxy-L-prolyl-4,5-dihydroxy-L-isoleucyl-6-hydroxy-2-mercapto-L-tryptophyl-glycyl-L-isoleucyl-glycyl-Lcysteinyl) cyclo(4  $\rightarrow$  8)-S-oxide. The octapeptide ring is bridged through the sulfur atom of the sulfoxide form of cysteine to the 2 position of the indole ring. The resulting bicyclic structure contains two 18-membered rings.

Crystals were grown by slow evaporation from a 95% eth-